Fungal Genetics Reports

Volume 27

Article 10

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Recommended Citation

Russell, P. J., and C.B. Perry (1980) "Cellobiose-induced B-galactosidase and B-glucosidase activities of Neurospora crassa.," *Fungal Genetics Reports*: Vol. 27, Article 10. https://doi.org/10.4148/ 1941-4765.1673

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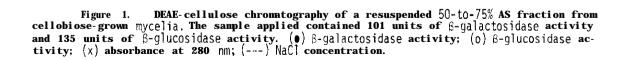
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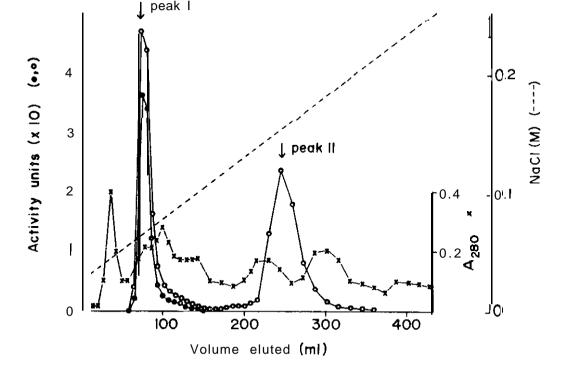
Abstract

Cellobiose-induced B-galactosidase and B-glucosidase activities of Neurospora crassa.

Russell, P. J. and C. B. Perry.Three β -galactosidase (β -D-galactoside galacto-
hydrolase: EC 3.2.1.23) activities have been shown in
wild type Neurospora. One of them is induced by
D-cellobiose, has an optimum activity at pH 6 and is pre-
cipitated at 50-to-75% saturation with ammonium sulfate
(AS) (Perry and Lester 1973 Biochem Biophys. Res.Commun.
54: 1476). Since D-cellobiose is an effective inducer
for β -glucosidases, "cellobiase," has optimum activity at
pH 6 form of β -galactosidase has any 3-glucosidase activity.

Beta-galactosidase and β -glucosidase activities were determined by use of the chromogenic substrates, q-nitrophenyl-3-D-galactopyranoside (ONPG) and p-nitrophenyl- β -D-glucopyranoside (PNPG). These and the other procedures used in this study (e.g. growth conditions, medium, cellobiose concentration, etc.), were described by Perry and Lester (1973 Biochem Biophys. Res. Commun. 54: 1476). A unit of enzyme activity is that amount which releases 1.0 µmOle of ONP or PNP per hour at 37°C under the assay conditions. For ion exchange Chromatography, diethylaminoethyl (DEAE) cellulose (Cellex D), WaS equilibrated with 0.01 M potassium phosphate, pH 6.8 containing 0.001 M EDTA, and poured into a 2.2 x 15-cm column (bed volume, 50 ml). Anmonium sulfate (AS) fractions were applied in volume of less than 5 ml and elution was carried out with a linear NaCl gradient (0.025 to 0.25 M) mde in 0.01 M potassium phosphate buffer.





When extracts of cellobiose-grown Mycelia were assayed at pH 6, both β -galactosidase and β -glucosidase activities were detected. The extracts were then subjected to AS fractionation. Very little of the enzyme activities (1% of the β -galactosidase and 4% of the β -galactosidase present in the crude extract) was precipitated by 0.50% AS; whereas, much more (75% of the β -galactosidase and 40% of the β -galucosidase) was precipitated by 50-75% AS. The 50.75% AS-fraction was then separated by chromatography on DEAE-cellulose. The fractions obtained were assayed for both activities (see Fig. 1). β -glucosidase activity appeared in two well-defined peaks (I and II), while β -galactosidase activity appeared in only one peak, which closely coincided with peak I of the β -glucosidase activity. There was, esentially, total recovery of both activities applied to the column. When activities were normalized to the peak maxim, the β -galactosidase peak and the β -glucosidase peak I were superimposable.

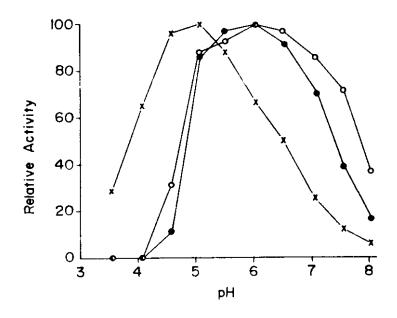


Figure 2. Effect of pH on DEAE-cellulose peak-! β -galactosidase activity (0), peak-1 β -glucosidase activity (o), and peak-II β -glucosidase activity (x). The number of units used were: peak-1 β -galactosidase, 1.30; peak-1 β -glucosidase, 1.30; and peak-11 β -glucosidase, 1.84. Activities were measured at pH optima.

Fractions corresponding to the two peaks were used to characterize further the β -galactosidase and β -glucosidase enzyme activities. Effects of pH on these activities are shown in Fig. 2. The pH optima of 5 and 6 for peak-II and peak-1 β -glucosidase activities are similar to the optima reported for $ary]-\beta-g]ucosidase$ (Mahadevan and Eberhart **1964** Arch. Bjochem. Biophys. 108: 22) and cellobiase (Eberhart and Beck 1970), respectively. The peak-1 β -glucosidase and β -galactosidase cannot be distinguished by the effect of pH upon their activities nor by the use of inhibitors. D-cellobjose or mercuric chloride. Finally, the peak-1 activities were tested for their thermal stabilities. At 50° C, β -galactosidase and β -glucosidase activities showed very similar kinetics of inactivation, with half-lives of 3 to 4 min.

In summary, these data show that the cellobiose-induced, pH 6- β -galactosidase of Neurospora is associated with a β -glucosidase activity, possibly cellobiase. It is not known at this time whether this is because Neurospora possesses a single enzyme activity that has dual specificity for β -galactoside and β -glucoside substrates, or whether there are two separate enzymes that copurify in the procedures used.

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