Fungal Genetics Reports

Volume 14

Article 25

Agar gel electrophoresis of amylases

D. N. Fass

Follow this and additional works at: https://newprairiepress.org/fgr



This work is licensed under a Creative Commons Attribution-Share Alike 4.0 License.

Recommended Citation

Fass, D. N. (1969) "Agar gel electrophoresis of amylases," *Fungal Genetics Reports*: Vol. 14, Article 25. https://doi.org/10.4148/1941-4765.2056

This Enzyme Methodology is brought to you for free and open access by New Prairie Press. It has been accepted for inclusion in Fungal Genetics Reports by an authorized administrator of New Prairie Press. For more information, please contact cads@k-state.edu.

Agar gel electrophoresis of amylases

Abstract

Agar gel electrophoresis of amylases

Fass, D. N. Agar get electrophoresis

of Neurospora amylases.

The procedure of Kikkawa (1963 Ann. Rep. Scient. Works, Fac. Sci. Osaka Univ. 11:41) previously used for the production of zymograms of Drosophila amylase hos been modified for use with <u>Neurospora crassa</u>. A gloss plate <u>ca</u>. 13 X 10 cm is frosted on one ride by grinding with household cleanser (Comet). The electro-

phoretic medium consists of 2% Difco Purified Agar in 0.01 M citric acid buffered to pH 5.0 with NaOH. This some buffer is used in the electrode reservoirs. A layer of tempered agar 1 mm thick is pipetted onto the frosted side of the prewarmed plate and is evenly with the ride of a pipette. Whatman #2 filter paper strips 1 x 0. 15 cm soaked with enzyme ore applied too line 3 cm from a long edge of the plate. The enzyme is absorbed for 10 min, after which the strips are removed. The plate is connected to the reservoirs by double thicknesses of Whatman #1 filter paper. A potential gradient of 40V/ccm is applied and maintained for 2 hours. The amylases will migrate to the cathode.

The plate is then immersed in a 4 mg/ml soluble starch solution for 15 min, followed by a brief water rinse. Digestion of the starch is allowed to occur for 20 min in a 37°C incubator, after which the agar is stained in a solution of 0.3% Kl = 0.03% 12. Two types of bonds will Lx visible; clear (against dork blue) and faint pink. The former ore the Y-amylases (gluc-amylases) and the latter are the a-amylaser.

Enzyme may be obtained by growing most strains for 8 days in Vogel's salts plus 2% SUCFOSE with necessary supplements in stationary culture. The medium is decanted and the pads are washed for 1 hr in Vogel's salts. This glso is decanted and Vogel's salts plus 1% maltase is added to the pad. After 24 hrs of shaking at 25°C, easily detectable quantities of enzyme will be present in the medium. This medium should be concentrated 20-50 fold by dialysis against air or sucrose before electrophoresis. Strain inos 89601a (FGSC#498), grown for 4 days with shaking in Vogel's salts + 1% sucrose and inosito, will produce sufficient enzyme in the medium for electrophoresis without concentration. Strain inos 89601A (FGSC#497) d oes not produce elevated levels of amylgse (H. G. Gratzner, person.1 commun.).

This work supported in part by the NSF and NIH Training Grant in Genetics (TO1-GMO1316) to Florida State University. = -Genetics Laboratories, Deportment of Biological Science, Florida State University, Tallahassee, Florida 32306.