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Abstract

We report the isolation and identification of a gene encoding GMP synthetase from a *Neurospora crassa* cDNA library. Phage infection of the purine-requiring *Escherichia coli* strain SØ3834 using the NO₃-induced cDNA phage library from the Fungal Genetics Stock Center resulted in colonies able to grow on minimal media with no added purine source. A plasmid, termed pGMPS1, was isolated from one of these colonies and shown to reproducibly support growth of strain SØ3834 in the absence of purines in the media. Identification of this gene as one encoding GMP synthetase is confirmed by DNA sequencing and comparison to the known *guaA* gene from yeast.

Identification of a gene encoding GMP synthetase from a *Neurospora crassa* cDNA library by bacterial complementation

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We report the isolation and identification of a gene encoding GMP synthetase from a *Neurospora crassa* cDNA library. Phage infection of the purine-requiring *Escherichia coli* strain SØ3834 using the NO₃⁻ induced cDNA phage library from the Fungal Genetics Stock Center resulted in colonies able to grow on minimal media with no added purine source. A plasmid, termed pGMPS1, was isolated from one of these colonies and shown to reproducibly support growth of strain SØ3834 in the absence of purines in the media. Identification of this gene as one encoding GMP synthetase is confirmed by DNA sequencing and comparison to the known *guaA* gene from yeast.

Nucleotide biosynthetic enzymes are common targets for anti-nucleic acid drug therapy, and deficiencies of the corresponding genes in humans are implicated in a number of inherited metabolic disorders. GMP synthetase (EC 6.3.5.2) has implications in anticancer and immunosuppressive therapies; additionally, this enzyme is used industrially in the semi-enzymatic synthesis of GMP (Fujio *et al.* 1997, *Bioscience, Biotechnology, and Biochemistry* 61: 840-845). The enzyme is a member of the amidotransferase family, which utilizes glutamine hydrolysis as a source of ammonia for a subsequent enzymatic step, and catalyzes the replacement of O₂ in XMP with an amino group to yield GMP.

To extend the collection of GMP synthetase genes from different organisms, we attempted isolation of the gene from *Neurospora crassa* using bacterial complementation. The auxotrophic *Escherichia coli* strain, SØ3834 (Chang *et al.* 1991, *Biochemistry* 30: 2273-2280), lacks both genes for GMP synthetase (*guaA*) and adenosine deaminase (*add*), and requires a purine source that can be metabolized to GMP for growth. In *guaA*⁻ *add*⁻ strains, the purine source can be 2,6-diaminopurine, since this compound can be enzymatically ribosylated and subsequently deaminated by adenosine deaminase to guanosine, but in SØ3834, 2,6-diaminopurine is incapable of providing the purine requirement. Thus, colonies in the complementation screen that arise on media containing 2,6-diaminopurine could contain either the *guaA* or *add* genes, but those that arise on purine-free media could only contain *guaA*.

From colonies arising in a complementation screen of about 10⁵ phage [NO₃⁻ induced cDNA phage library, Fungal Genetics Stock Center; *Fungal Genetics Newsletter*, 45 (Supplement), 8], one colony yielded plasmid DNA from an alkaline lysis preparation that, when reinserted into the host strain, provided the ability to grow on purine-free media. This plasmid was presumed to contain the GMP synthetase gene and was designated pGMPS1. Sequencing of the inserted region within the vector was carried out at the Automated DNA Sequencing Facility, University of North Carolina, Chapel Hill, using the M13 (-20) primer and primers designed from newly acquired sequence information. An open reading frame of 1629 nucleotides was identified; when translated into the amino acid sequence, a high degree of similarity was found with the yeast GMP synthetase gene (duJardin *et al.* 1994, *Gene* 139:127-132). Of the 543 amino acids from the deduced sequence, 354 (65%) have identical counterparts in the yeast sequence (Figure 1). The high degree of sequence similarity with the yeast GMP synthetase gene and the ability of the gene to support growth of *E. coli* strain SØ3834 in the absence of purines confirm the identification of a GMP synthetase gene from an *N. crassa* library.

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Figure 1. Amino acid sequence comparison of GMP synthetases. Yeast sequence is shown in light type, sequence from *N. crassa* is bold. Vertical dashes between sequences indicate identical amino acids.

MAA	GEQV	SNMFDTILVL	DFGSQYSHLI	TRRLREFNIY	AEMLPCTQKI	
MAAATLGEVP	TKAFDTILTL	DFGSQYTHLI	TRRMRELNIL	SEMLPCTTKI		
SELGWTPKGV	ILSGGPYSVY	AEDAPHVDHA	IFDLNVPILG	ICYGMQELAW	97	
ADLDYKPKGV	ILSGGPYSVY	EDGAPHVDPA	VFELGVPILG	ICYGMQEIWA	100	
INGKQVGRG	DKREYGPATL	KVI	DDSNS	LFKGMNDS	TVWMSHGDK	
RASPENVIAG	VHREYGHNSL	KALKGDDAHV	DRLFAGLEDG	MRVWMSHGDK		
LHGLPTGYKT	IATSDNSPYC	GIVHETKPIY	GIQFHPEVTH	STQGKTLLKN	191	
LGALPEGFHT	VAVSDNSEYA	AIAHKTKPIY	GLQFHPEVTH	SQNGTQLLKN	200	
FAVDLCHAKQ	NWTMENFIDT	EINRIRKLVG	PTAEVIGAVS	GGVDSTVASK		
FAVDICGCAQ	NWTMARFLDQ	EIARIRDLVG	PEGQVLGAVS	GGVDSTVAAK		
LMTEAIGDRF	HAILVDNGVL	RLNEAANVKK	TLVEGLGINL	MVVDASEEFL	291	
LMKEAIGDRF	WAVLVNNGVM	RLDECEQVER	DLKQHLGINL	TVIDASKDFL	300	
SKLKGVTDPK	KKRKIIGNTF	IHFEREAEK	IKP	KDGK	EIQFLLQG	
EGLKGLHDPE	QKRKFIGGKF	IDVFEAEAQK	IEEAAKSGK	GTKIGFFLQG		
TLYPDVIESI	SFKGPSQTIK	THHNVGGLLE	NMK	LKL	IEPLRELFKD	382
TLYPDVIESL	SFKGPSATIK	THHNVGGLPE	RMTNGQGLQL	IEPLRSLYKD	400	
EVRHLGELLG	IPHDLVWRHP	FPGPGIAIRV	LGEVTKEQVE	IARKADNIYI		
EVRELGRITL	IHEELVMRHP	FPGPGIAVRI	LGEVTEEKVR	IARQADHIFI		
EEIKKAGLYN	QISQAFACLL	PVKSVGVMGD	QRTYDQVIAL	RAIETTDGMT	482	
SEIRKAGLYD	QISQAYAAVD	PSRAVGVMGD	KRVYGYIIIL	RAVTTTDMGT	500	
ADWFFFEHSF	LKKVASRIVN	EVDGVARVTY	DITSKPPATV	EWE	525	
AEAFNFPWDF	LQRMNRIVN	EVNGVCRVTY	DITSKPPGTI	ELE	543	